



## Biocidal spectrum of a rice field cyanobacterium *Nostoc* sp.

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### Abstract

The antimicrobial efficacy of hexane, dichloromethane and ethyl acetate extracts of a rice-field cyanobacterium, *Nostoc* sp., were evaluated against cyanobacteria and phytopathogenic fungi. The maximum production of biocidal compounds was observed in cultures grown for 20 days under optimized conditions (phosphate =1.4 mg/l; light intensity ≈ 3000-4000 lux). Fractionation of the extracts showed six spots on silica-gel-coated plates. Hexane extract was the most potent biocide, showing marked (35-42%) reduction in the growth of fungi, but no negative influences on seed germination or seedling growth of wheat, rice and mung bean, emphasizing its suitability for use in agriculture.

**Keywords:** Antifungal; extract; inhibition zone; metabolite; *Pythium debaryanum*; *Rhizoctonia solani*.

### Introduction

Cyanobacteria constitute a heterogeneous group of phototrophic prokaryotes with characteristics in common to both bacteria and algae. They are often referred to as 'biofactories' due to their unique biosynthetic potential to produce wide range of compounds e.g enzyme, hormones, pigments, proteins, fatty acids/lipids, etc (Prasanna *et al.* 2010). They are distributed in a wide range of aquatic and terrestrial habitats owing to their high adaptive ability under diverse environmental conditions. Some species of cyanobacteria can proliferate in aquatic environments, leading to a significant deterioration of water quality- a phenomenon known as "water bloom". These water blooms are of tremendous biological significance because they have proved excellent sources of exciting molecules with immunomodulatory, bioregulatory and therapeutic potential (Namikoshi & Rinehart 1996, Prasanna & Jaiswal 2006, Jaiswal *et al.* 2007). However, increasing evidence indicates that besides these bloom forming strains, cyanobacteria colonizing other habitats may also produce wide variety of low molecular weight secondary metabolites with biological activities (Ghasemi *et al.* 2003; Jaiswal *et al.* 2005; Jaiswal *et al.* 2008; Prasanna *et al.* 2010).

The range of bioactivities shown by cyanobacterial compounds is very diverse and include antimicrobial, antiviral, immuno-modulatory, insecticidal and antioxidant activities, which can be agriculturally useful. They also serve as allelochemicals, providing a competitive advantage to the producer and contributing to the dominance of a strain in a particular habitat. A number of compounds have been isolated that exhibit biocidal activity against specific agriculturally important pathogens (fungi, nematodes) (Pushparaj *et al.* 2000, Nagle & Wedges 2002, Volk & Fulkert 2006, Prasanna *et al.* 2009, Chaudhary *et al.* 2010, Manjunath *et al.* 2010). Therefore cyanobacteria offer an effective soil-friendly alternative to agrochemicals for controlling plant pathogens while maintaining soil fertility and environmental safety. Cyanobacteria from local habitats can be a good source of potential biologically active substances responsible for the controlling the growth of local pathogenic microorganisms (Dukare *et al.* 2011). With this in mind, the current investigation was aimed at evaluating the biocidal spectrum of a rice-field cyanobacterium, *Nostoc* sp., and its potential as a biocontrol agent.

## Materials & Methods

*Nostoc* sp. used in this investigation was isolated from a rice field grown at the Indian Agricultural Research Institute (Jaiswal *et al.* 2005). Two unicellular cyanobacterial strains (*Synechococcus* sp. and *Synechocystis* sp.) were used as test organisms. All these strains were grown and maintained in BG11 medium and incubated at  $27 \pm 1$  °C, 3000 lux light intensity and 16:8 L:D cycles. The fungal strains (*Pythium debaryanum*, *Fusarium oxysporum*, *Rhizoctonia solani*, *Rhizoctonia bataticola* and *Alternaria solani*) used for evaluating fungicidal activity were obtained from the Division of Plant Pathology, IARI, New Delhi. The cultures were maintained on potato dextrose agar medium. The fungal strains were grown and maintained at  $28 \pm 2$  °C in a BOD incubator.

In order to study the effect of different physicochemical factors on algicidal activity, the basal medium was supplemented with different levels of P (half and twice the levels of the BG-11 medium). The effect of light intensities (1500-6000 lux) and temperature (20-40 °C) was also evaluated.

The axenized isolate of *Nostoc* sp. was set for mass multiplication in 3L flasks and incubated under optimized conditions for maximum biocidal activity. The cultures were harvested in the exponential phase of growth by pelleting at high speed (8000 rpm) for 20 minutes. The cell free extracts were pooled for the extraction of secondary metabolites.

The culture filtrates were partitioned with non-polar (n-hexane), polar (dichloromethane) and highly polar (ethyl acetate) solvents sequentially to extract all the compounds present in culture filtrates. The organic phase was then separated from the aqueous phase and dried over anhydrous sodium sulphate. The extracts were then subjected to rotary evaporation under pressure. The solid extract was dissolved in acetone and stored at 4 °C until further analyses.

The bioactivity of the cyanobacterial filtrate, crude extracts and the partially purified compounds were tested on a lawn of two unicellular cyanobacteria (*Synechococcus* sp. and *Synechocystis* sp.) and five phytopathogenic fungi by the disc-diffusion technique (Jaiswal *et al.* 2005). The diameter of the inhibition zone was taken as the measure of toxicity. The percent inhibition of the fungal strain was also calculated by the food-poisoning technique and measuring the colony size of the fungal strain.

The inhibitory effect of the concentrated extracts was examined in agar-based media using 96-well ELISA plates based on the methodology developed by Prasanna *et al.* (2006). After rotary evaporation, extracts were weighed and dissolved in the desired volume of acetone and a series of different dilutions prepared. Five replicates of each concentration were placed in sterilized medium (BG-11 and potato dextrose agar) in the wells of the ELISA plates. Each well was inoculated with the test organism (cyanobacterium/fungus) and incubated under optimal growth conditions along with suitable controls.

The effect of culture filtrate/crude extract of the cyanobacterium was tested on seed germination, length of coleoptile and radicle of wheat (var HD2689), rice (var Pusa Basmati 1) and mung bean (var Pusa Ratna). Seeds were surface-sterilized using 0.1% mercuric chloride and washed thoroughly with sterile water. They were imbibed overnight in sterile water in the dark at room temperature and kept for germination on water agar (2% agar in distilled water containing minimum inhibitory concentration levels of cyanobacterial extracts). The plates were incubated in the dark at  $25 \pm 2$  °C. The percentage germination of the seeds was calculated after 24 h of incubation, and the length of the coleoptile and radicle measured after 48 h (Jaiswal *et al.* 2010).

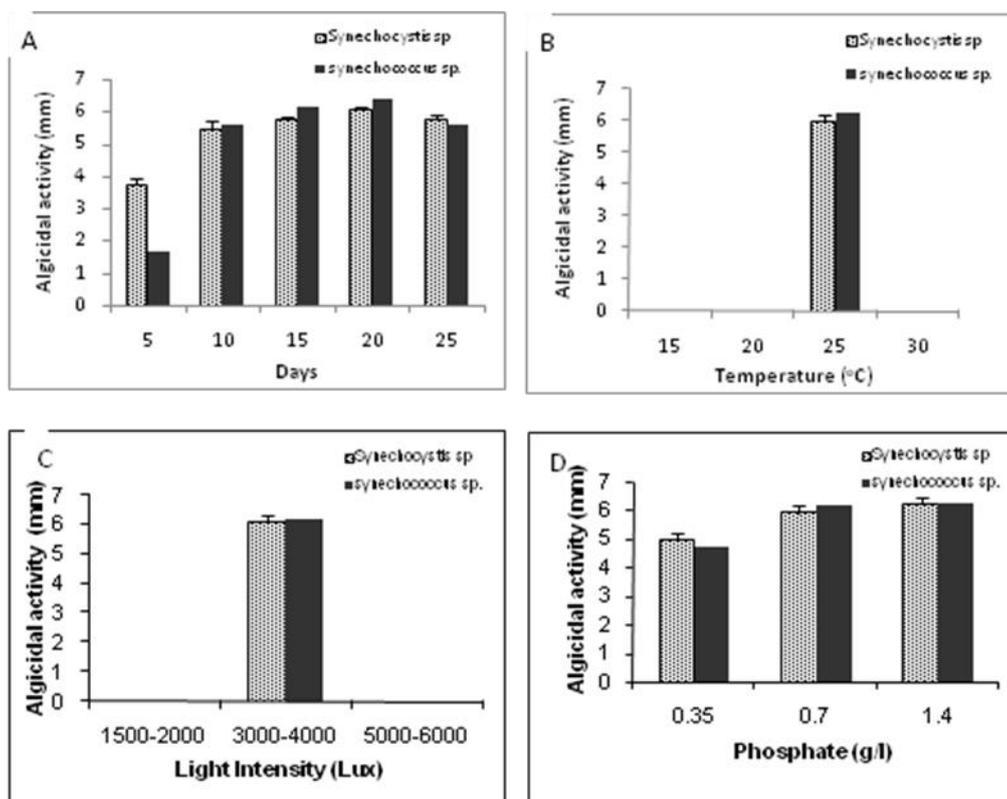
Thin-layer chromatography of the samples separated the compounds present in the *Nostoc* extracts (Jaiswal *et al.* 2010). Silica-gel-coated plates (0.15 mm thick) activated in a hot air oven at 120 °C were used. The crude extract dissolved in acetone was spotted on the TLC

plates and then passed through different solvent systems to achieve maximum separation of the constituent compounds.

Statistical analysis was carried out using Microsoft Excel. At least three independent experiments were conducted and readings were taken in triplicate for each.

## Results

The culture filtrate of *Nostoc* showed algicidal activity (Jaiswal *et al.* 2005), which was significantly affected by culture age. Time-course studies indicated that 5-d-old cultures (early exponential phase) exhibited very little algicidal activity, but gradual enhancement was observed with increasing culture age (Figure 1A). The maximum algicidal activity was recorded with 20-d-old cultures, which were 3.8- (*Synechococcus*) and 2-fold (*Synechocystis*) higher than 5-d-old culture. Further ageing (to 25 days) led to a slight decline in biocidal activity.

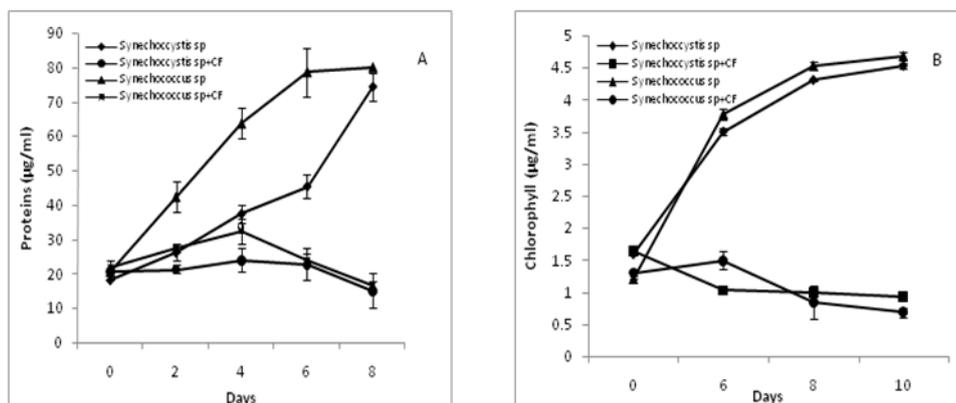


**Figure 1:** Effect of incubation (A), temperature (B), light intensity (C) and phosphate levels (D) on the algicidal activity of *Nostoc* sp. (measured as diameter of the inhibition zone).

A shift in the temperature from the optimal range ( $25 \pm 2$  °C) resulted in complete loss of biocidal activity (Figure 1B). Similarly, a change in light intensity from the optimal level (3000- 4000 lux) also resulted in loss of algicidal activity (Figure 1C). The highest algicidal values were recorded when the phosphate concentration was doubled (1.4 g/l) as compared to basal level in BG-11 medium, but there were significant differences between the two test organisms. The algicidal activity at lower levels of P (0.35 g/l) was substantially reduced (Figure 1D).

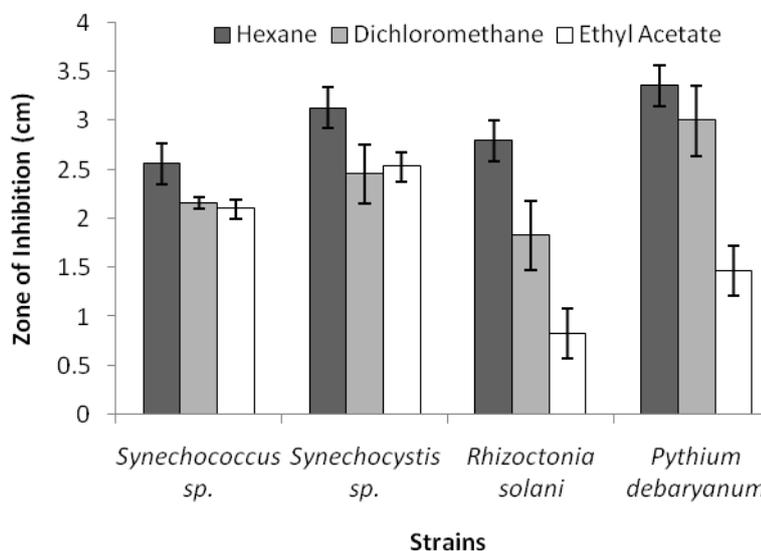
The addition of culture filtrate @ 10% (v/v) of *Nostoc* sp. led to a 1.5- (*Synechocystis*) and 1.2-fold (*Synechococcus*) lowering of protein content on the 2<sup>nd</sup> day of incubation (Figure

2A). It reduced much further by the end of 8<sup>th</sup> day. The chlorophyll content was also strongly affected by the culture filtrate of *Nostoc* sp. (Figure 2B), 4.9- (*Synechocystis*) and 6.9-fold (*Synechococcus*) lower than the control on the 8th day of incubation.



**Figure 2:** Effect of culture filtrate (CF) of *Nostoc* sp. on protein (A) and chlorophyll (B) content of *Synechococcus* and *Synechocystis* sp.

The crude extracts of cyanobacteria (grown under optimized conditions) using n-hexane, dichloromethane and ethyl acetate were tested for biocidal activity on the two unicellular cyanobacteria and five phytopathogenic fungi. Biocidal activity was recorded in all the extracts with varied sensitivity (Figure 3). The hexane extract had maximum biocidal (algicidal and fungicidal) activity in terms of the zone of inhibition. Differences in algicidal activity by the dichloromethane and ethyl acetate extracts were not significant, but the former was observed to be more potent in showing fungicidal activity.



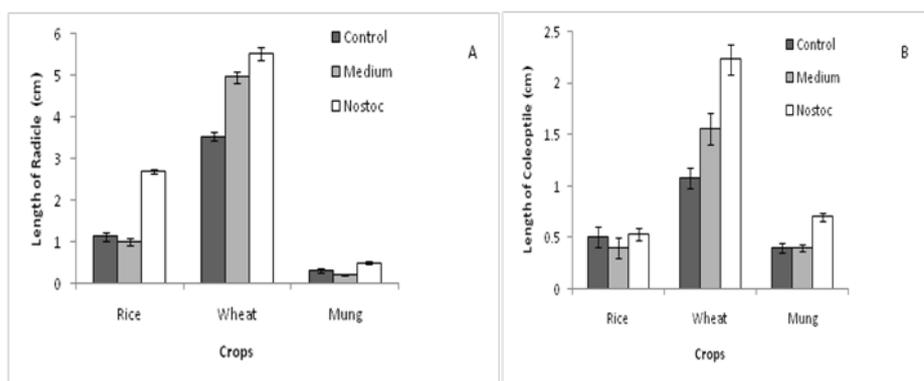
**Figure 3:** Biocidal activity (measured as zone of inhibition) of crude extract of *Nostoc* sp. against two unicellular cyanobacteria (*Synechococcus* and *Synechocystis* sp.) and two phytopathogenic fungi (*R. solani* and *P. debaryanum*).

The crude extract also showed significant reductions in growth of the cyanobacterial and fungal strains (Table 1). The maximum inhibition was recorded with the hexane extract. Among the two fungal strains, *P. debaryanum* exhibited a 42% reduction, while among the two cyanobacterial strains *Synechocystis* sp. was found to be more sensitive to the extract.

Test organism	*Reduction in growth (%)		
	n-hexane	dichloromethane	ethyl acetate
<i>Synechococcus</i> sp.	32.0	27.0	26.3
<i>Synechocystis</i> sp.	39.1	30.8	31.6
<i>Rhizoctonia solani</i>	35.0	22.8	10.3
<i>Pythium debaryanum</i>	42.0	37.5	18.4

**Table 1:** Effect of cyanobacterial extract (*Nostoc* sp.) on the percent reduction in growth of test organisms.

The minimum inhibitory concentration required for the algicidal and fungicidal activity was calculated by serial dilution of the concentrated extract: for algicidal activity this was found to be 31.2 ppm, while it was 62.5 ppm for fungicidal activity.



**Figure 4:** Effect of cyanobacterial culture filtrate on length of the radicle (A) and coleoptile (B) of rice, wheat and mung seedlings.

The effect of the culture filtrate/extract on important crop plants was studied. Two concentrations of the extract were used - 31.25 ppm and 62.5 ppm (based on the minimum inhibitory concentration). The lower concentration had no effects on germination of wheat, mung and rice seeds (data not shown). The culture filtrate enhanced radicle and coleoptile lengths for all three crop seedlings (Figure 4). The extracts had no negative influences, except slight reduction in lengths for mung bean seedlings at the 62.5 ppm concentration (Table 2).

Treatments	length (cm)					
	wheat		rice		mung bean	
	coleoptile	radicle	coleoptile	radicle	coleoptile	radicle
Control (water)	1.08 ± 0.05	3.53 ± 0.58	0.50 ± 0.02	1.13 ± 0.10	0.40 ± 0.10	0.30 ± 0.06
Control (acetone)	1.04 ± 0.03	3.53 ± 0.36	0.50 ± 0.05	1.12 ± 0.00	0.40 ± 0.00	0.30 ± 0.10
31.25 ppm	1.10 ± 0.07	3.70 ± 0.06	0.50 ± 0.08	1.15 ± 0.10	0.40 ± 0.10	0.30 ± 0.10
62.50 ppm	1.08 ± 0.10	3.65 ± 0.20	0.50 ± 0.02	1.15 ± 0.03	0.36 ± 0.03	0.28 ± 0.11

**Table 2:** Effect of different concentrations of cyanobacterial extract on the length of coleoptile and radicle of selected seedlings.

Thin-layer chromatography fractionation of crude cyanobacterial extract showed the presence of 2 (n-hexane), 5 (dichloromethane) and 3 (ethyl acetate) spots (Table 3) when n-hexane: benzene (1:1) was used as the solvent system. Fraction A with R<sub>f</sub> value 0.07 was common to all solvents, while fraction B was common to hexane and dichloromethane, and

fraction C to dichloromethane and ethyl acetate solvents. Fractions D (ethyl acetate), E and F (dichloromethane) were unique to single solvents.

Solvent	bands	Rf values					
		A	B	C	D	E	F
n-Hexane	2	0.07	0.16	-	-	-	-
Dichloromethane	5	0.07	0.16	0.22	-	0.60	0.94
Ethyl acetate	3	0.07	-	0.22	0.29	-	-

**Table 3:** Comparative rate of flow of different fractions of crude extract of *Nostoc* sp. using thin layer chromatography.

The extracted metabolite (n-hexane and dichloromethane extracts) was partially purified using preparatory thin layer chromatography and fraction A was found to exhibit the maximum algicidal activity, followed by fractions B and E; maximum fungicidal activity was recorded for fraction B followed by fraction A. Other fractions did not show any fungicidal activity.

## Discussion

Cyanobacteria, a large group of structurally diverse, ecologically adaptive, Gram negative prokaryotes, have long been known for their potential use in agriculture as biofertilizer, soil conditioner and plant growth regulation due to their ability to secrete a number of compounds such as polysaccharides, peptides, etc, during growth. They have recently drawn the attention of biologists worldwide for their rich source of novel bioactive compounds with multifaceted applications (e.g. in agriculture, industry, pharmaceuticals). Studies have revealed that secondary metabolite production in cyanobacteria is significantly influenced by culture age. Most of the reports indicate that it is released into the medium in the late exponential phase after cell lysis (Ray & Bagchi 2001, Dias *et al.* 2002). In the current investigation with *Nostoc* sp., algicidal activity was detected, on the lawn of two unicellular cyanobacteria (*Synechococcus* and *Synechocystis* sp.) at all the stages of growth, increasing with time and with a maximum in the stationary phase culture. The correlation between culture age and activity is similar to earlier reports (Ray & Bagchi 2001, Dias *et al.* 2002). Volk (2007) also reported that the content of exo-metabolites of a cyanobacterium *Nostoc insulare* changes with growth phase; during the linear growth phase, a non-toxic metabolite was prevalent, while in the stationary phase the content of antimicrobial cytotoxic metabolite was enhanced.

The cyanobacterium in the current investigation showed loss of algicidal activity when grown in culture conditions (i.e light and temperature) other than the optimal range. There have been contradictory reports regarding the influence of light and temperature on the production of biocidal compounds. A few reports suggest that an increase in light intensity has a positive effect on toxin production (Utkilin & Gjolme 1992), while some others suggest that there is no significant effect (Wicks & Thiel 1990). Most of the earlier reports also suggest that toxin production by cyanobacteria is less at the upper and lower limits of the temperature optima for the species (Ressom *et al.* 1994, Jaiswal *et al.* 2007, Jaiswal *et al.* 2008), which agrees with the results obtained in the current investigation.

In order to study the effect of algicidal compound on the test organism, the protein and chlorophyll content of two unicellular cyanobacteria were estimated. The addition of culture filtrate brought about a significant reduction in the protein and chlorophyll content by the 2<sup>nd</sup> day of incubation, indicating that the metabolite may affect some of the primary metabolic processes. Bagchi & Ray (2001) also reported that the cyanobacterial extract of *Oscillatoria* sp. showed PSII-inhibiting activity, besides algicidal activity.

In order to understand the polar/nonpolar nature of the biocidal metabolite, the culture filtrate was partitioned using different solvents. Maximum activity was observed in hexane extract, indicating the non-polar nature of the active metabolite. TLC fractionation of the extracts resolved into a number of spots, indicating the presence of more than one compound in each extract, further confirmed by presence of a number of peaks in the GC analyses. Among the various fractions, algicidal activity was shown by fraction no. A, B and E in descending order of activity. The presence of a large number of metabolites with varying toxicity levels and host range has been reported in cyanobacteria (Jaki *et al.* 2000, Volk & Mundt 2007, Jaiswal *et al.* 2010).

The effect of cyanobacterial filtrate and crude extract on seed germination and growth of important crop plants revealed the positive influence of culture filtrate on seedling growth, which may be due to the presence of some growth-promoting substances in the cyanobacterial extract. Though we have not tested the cyanobacterial extract for the presence of amino acids, lipids or indole acetic acid, such biomolecules are not uncommon in cyanobacterial extracts (Kartikeyan *et al.* 2009). More importantly, the crude extract of the culture filtrate did not have any negative influence on seed germination and growth of seedlings, an important advantage for the potential use of extract as a fungicidal agent, because non-target biota are not affected.

Cyanobacteria have long been known for their potential for N<sub>2</sub> fixation in rice fields, and hence have been traditionally used as biofertilizer, with only few reports on the production of antimicrobial substances from rice-field cyanobacteria (Ghasemi *et al.* 2003). The rice rhizosphere is known to be the site of a large number of microorganisms belonging to diverse groups including cyanobacteria (Prasanna *et al.* 2009). The presence of biologically active substances may serve as a defence strategy against other co-occurring organisms such as bacteria, fungi and viruses. The current study highlights the potential of these cyanobacteria as biocontrol agents against various economically important plant pathogens, thus enhancing their value as inoculants.

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